Design of a Compound

Screening Collection

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In the Past...

- Scientists chose what molecules to make

- They tested the molecules for relevant activity
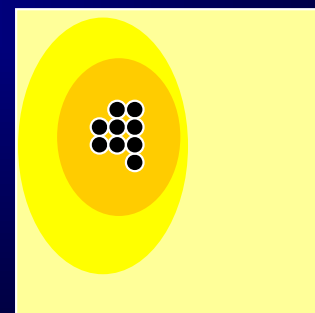
Now...

- We often screen a whole corporate collection
  - 10^5-10^6 compounds

- But we choose what's in the collection

- If the collection doesn't have the right molecules in it
  - we fail

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"Screen MORE"

- Everything'll be fine
- We'll find lots of hits
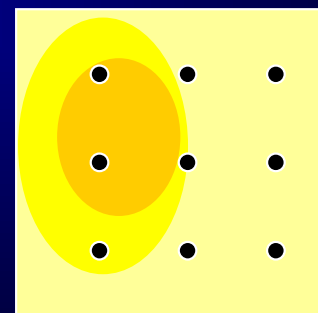

- Not borne out by our experience

How do I design a collection? - 1

- Pick the right kind of molecules
    - hits similar biological targets
    - computational (in-silico) model predicts activity at right kind of target for given class of molecules
    - exclude molecules that fail simple chemical or property filters known to be important for "drugs"

- FOCUS!

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How do I design a collection? - 2

- Cover all the options
- Pick as "diverse" a set of molecules as possible
- If there's an active region of chemical space, we should have it covered

- DIVERSE SELECTION
  - opposite extreme to focused selection

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Basic Idea of Our Model

- Relate biological similarity to chemical similarity
- Use a realistic objective
  - maximize number of lead series found in HTS
- Build a mathematical model on **minimal** assumptions

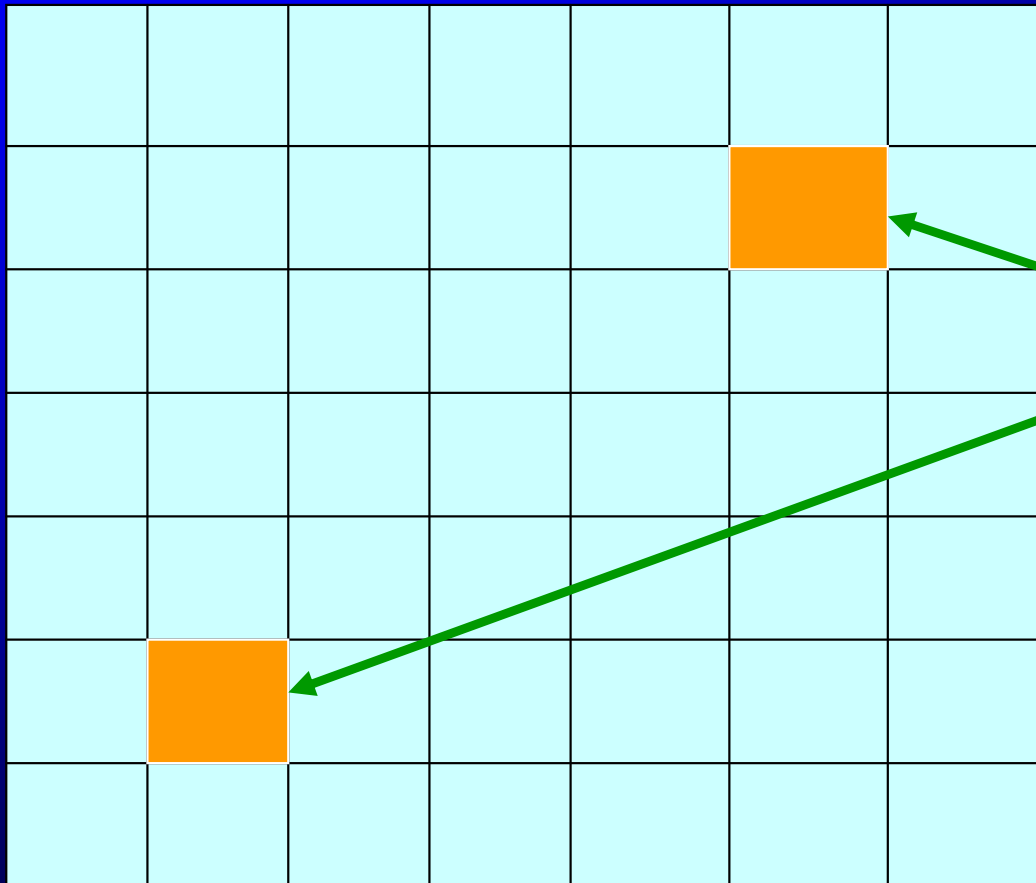$\Rightarrow$ How does our collection perform now in HTS?
  - relate this to our model

$\Rightarrow$ Learn what we need to make/purchase for HTS to find more leads

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A "simple" model

- Chemical space is clustered (partitioned)
  - there are various possible ways to do this

- For a given screen, each cluster i has

  - a probability π_i that it contains a lead
- If we sample a random compound from a cluster containing a lead, the compound has

  - a probability α_i that it shows up as a hit in the screen
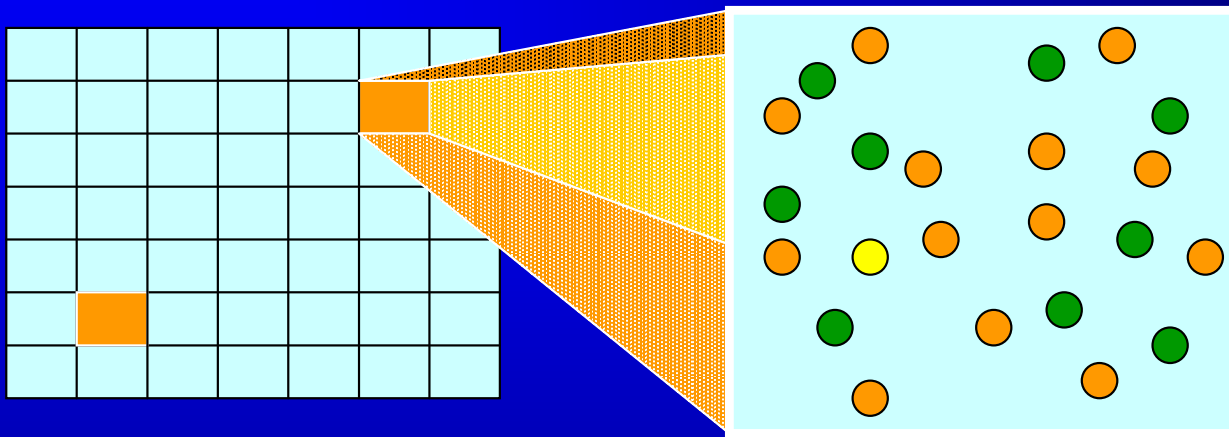- If we find a hit in the cluster, that's enough to get us to the lead

And in pictures...

clusters containing leads

$\pi_i = \text{Pr}(\text{box } i \text{ is orange})$

Hit
Non-Hit
Lead

$\alpha_i = \Pr(\text{dot is green})$

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Constrained Optimization Problem

- Suppose that we want to construct a screening collection of fixed size M

- To maximize expected number of lead series found we have to

$$
\textbf{(P)} \quad
\begin{aligned}
\text{Maximize} \quad & \sum_{i=1}^{p} \pi_i [1 - (1 - \alpha_i)^{N_i}] \\
& \qquad\qquad\qquad\qquad N_i \geq 0 \quad (i = 1, \ldots, p) \\
\text{subject to} \quad & \sum_{i=1}^{p} N_i = M
\end{aligned}
$$

Solution

$$N_i = \begin{cases} \dfrac{\ln \lambda - \ln \pi_i - \ln(-\ln(1-\alpha_i))}{\ln(1-\alpha_i)} & \text{whenever this is} \geq 0 \\[2em] 0 & \text{otherwise} \end{cases}$$
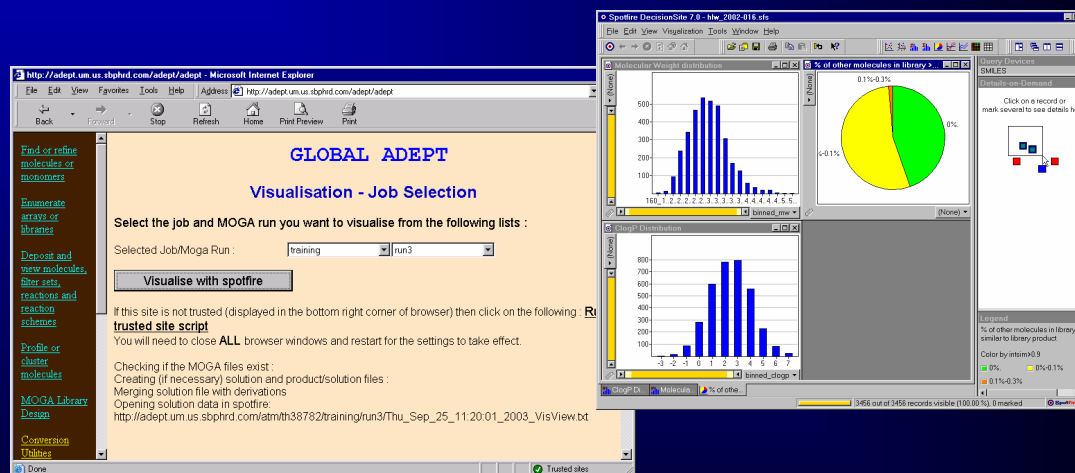
- If we know very little (α_i, π_i equal for all i)
  - select the same number from each cluster - **diversity solution**
- If e.g. we know some clusters are far more likely than others to contain leads for a target
  - select compounds only from these clusters - **focused solution** (filters)
- But we also have a solution for all the situations in between, where there is a balance between diversity and focus

Immediate Impact

- Improved "diversity" score

$$D(\{N_i\}_{i=1}^p) = \sum_{i=1}^{p}[1-(1-\alpha)^{N_i}]$$

- Use in assessing collections for acquisition

- We have integrated this score into our Multi-Objective Library Design Package

* Gillett et al., *J. Chem. Inf. Comp. Sci.* **2002**, *42*, 375-385.

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What value should α take?

- Determining a value of α is important. We can cluster molecules using a variety of methods.

- Fortunately, there is a recent paper from Abbott which answers this question

- In 115 HTS assays, with a TIGHT 2-D clustering, $\alpha \sim 0.3$
  - consistent: mostly varies between 0.2 and 0.4

- This agrees well with our experience

- In practice we use this (Taylor-Butina) clustering with radius 0.85 and using Daylight fingerprints

* Martin et al., *J. Med. Chem.* **2002**, *45*, 4350-4358.

- **A consistent value of α is necessary, irrespective of cluster**
- **Otherwise, very difficult to parameterise model accurately**

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The Rights of a Molecule

- Every molecule has the right to be treated equally
  - The probability of similar biological activity at similarity x should be the same, independent of bit density (or any other global properties)

- Our limited experience suggests larger molecules may be less likely than small molecules to be active using our 0.85-radius clustering
- Needs further exploration
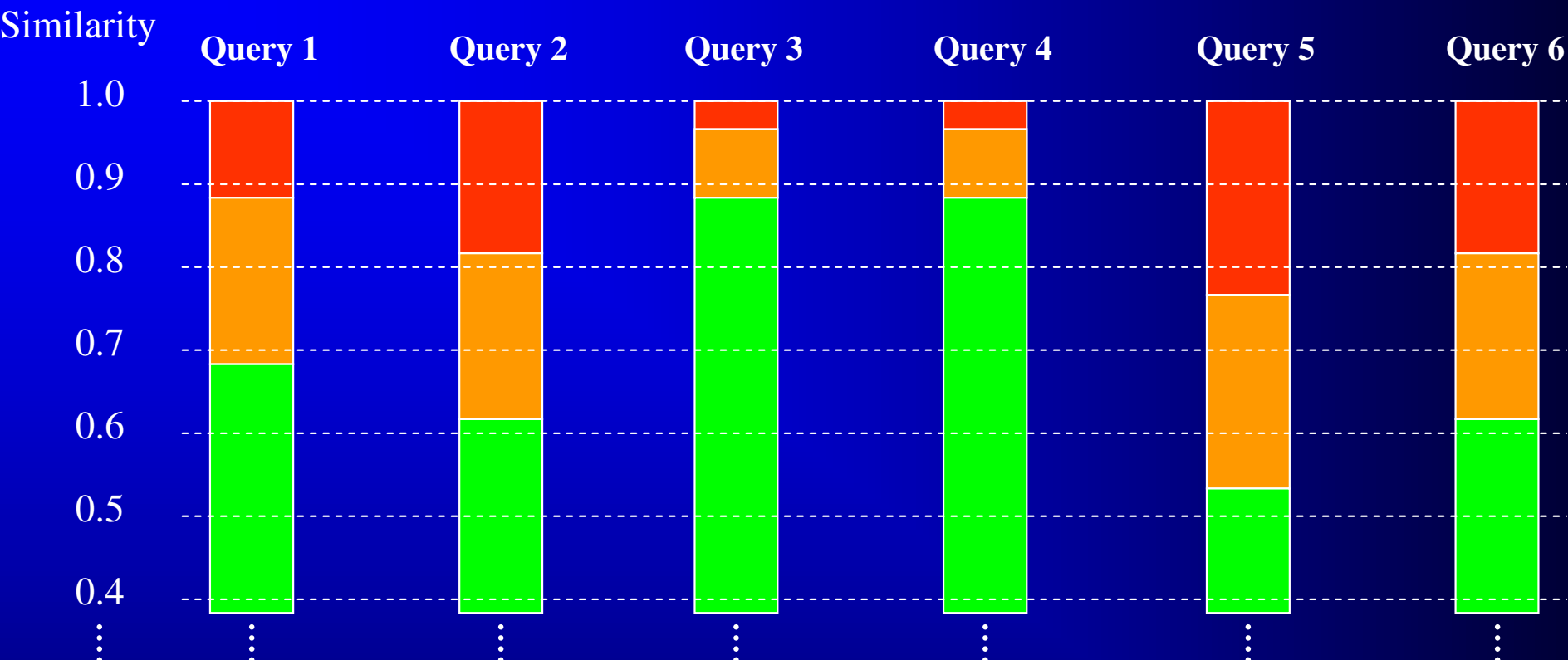 - But would we expect this to happen?

Recent papers: bit density vs similarity

– Flower: JCICS 48, 379-386 (1998)

– Fligner et al. Technometrics 44, 110-119 (2002)*

– Holliday et al. JCICS 43, 819-828 (2003)

– * In Fligner et al., they propose a simple random model.

  • Compare 2 molecules of same bit density:

  • Under model, expected Tanimoto similarity is approx $p/(2-p)$

    – where p is proportion of bits set

  • More dense bit strings

    ➢ higher Tanimoto similarity

But it doesn't just matter for my model!

- Papers were mainly concerned with dissimilarity problems
  - Easier to find low bit density compounds with near-zero similarity to existing compounds
    - Sequential dissimilarity-based selection bias

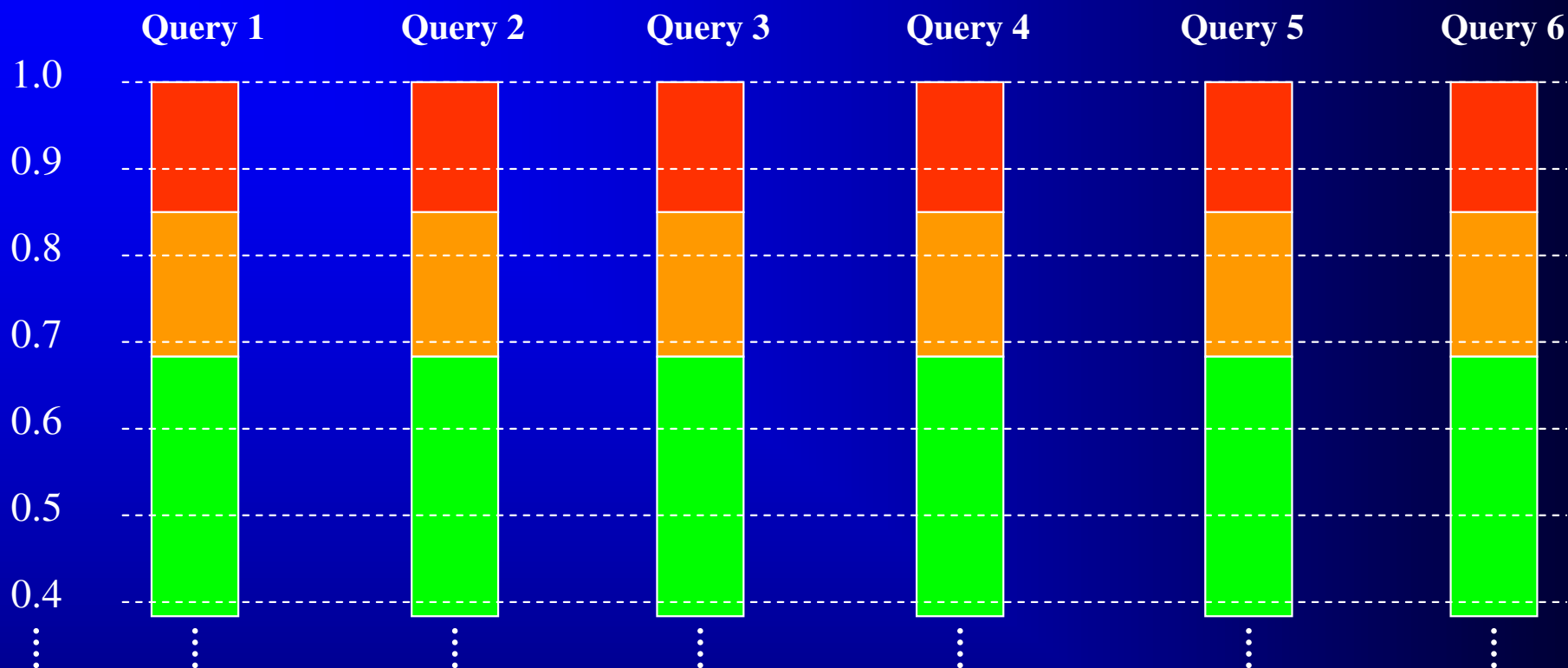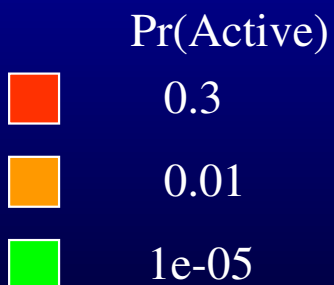- But consider similarity searching with multiple queries.

Life would be easier if…

- Finally of course
  - Use "the model" to work out which molecules to actually screen
  - It won't just be the top n if they're all highly similar to each other

Applications

- Compound acquisition
- Library design
- Strategic Decision-Making Tool
  - Resource allocation - what to buy, what to make.
  - What targets to screen
- Prioritisation of hits in virtual screening
  - Similarity searching
  - Pharmacophore searching?
  - Docking?
- Others?...

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Acknowledgements

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* Harper et al., *Combinatorial Chemistry and High Throughput Screening* **2004,** *7*, 63-70.

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